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Development of LymphoTrack[®] **Bioinformatics Methods**: **Clonality Testing, Somatic Hypermutation and Minimal Residual Disease**

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Introduction

- · Clonality assessment is an important diagnostic indicator for the malignant transformation of lymphoid cells
- Minimal Residual Disease (MRD) is a key prognostic indicator for determining efficacy of treatment
- . IGHV Somatic Hypermutation (SHM) is a naturally occurring process to increase B-cell receptor diversity, and serves as a positive prognostic factor in Chronic Lymphocytic Leukemia (CLL).
- Next-Generation Sequencing (NGS) assays for measuring these disease markers are becoming the cutting edge of diagnostic methods, but require the development of new bioinformatics tools to confidently analyze and report the results in a simple manner. We have developed the LymphoTrack[®] bioinformatics tools to analyze IGH, IGK and TRG genes on the Illumina[®] MiSeq[®] and Thermo Fisher Ion PGM* platforms, for the purpose of measuring clonality, *IGHV* SHM and MRD.

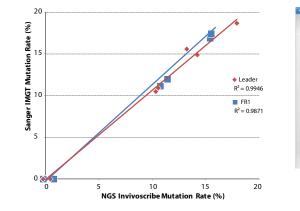


FIGURE 4. SHM CONCORDANCE

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97.82

97.82

97.82

97.82

97.82

97.82

97.82

97.38

97.82

LymphoTrack^{*} SHM calculations for IGHV FR1 and Leader closely match results generated by Sanger sequencing and IMGT mutation rate calculations.

FIGURE 5. MRD OUTPUT EXAMPLE Example output for MRD module. Results include the read count and

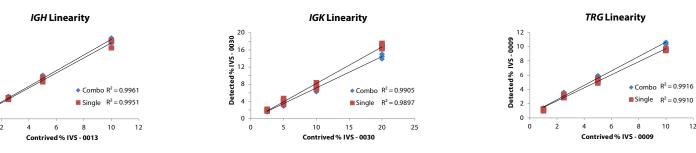


FIGURE 7. MULTIPLEXING CAPABILITY

LymphoTrack' Software is able to analyze samples mixed with all different target loci (IGH, IGK and TRG) even when sharing barcodes without a significant change in signal.

· LymphoTrack* bioinformatics analysis accomodates a wide variety of important diagnostic and * LymphoTrack* bioinformatics analysis also provides the resources necessary to delve deeper prognostic indicators. The straightforward presentation of results ensures an equally uncomplicated into the data by providing full sequence information and V-J assignment, allowing for an determination of clonality, SHM, and MRD. unprecedented level of granularity previously unattainable using capillary electrophoresis data.

Materials and Method

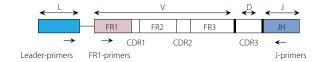


FIGURE 1. PRIMER DESIGN

LymphoTrack^{*} assays have been developed for both the MiSeq^{*} and PGM^{*} NGS platforms, for the IGH, IGK and TRG loci. Each assay employs a separate master mix to amplify the respective upstream V-region (or Leader) with the downstream J-region. Depicted is the IGH locus. The MiSeq* assays use 24 indices, allowing for parallel analysis of 22 samples (plus 2 controls). PGM* assays use 12 indices, allowing for parallel analysis of 10 samples (plus 2 controls).



FIGURE 2. BIOINFORMATICS WORKFLOW

GCCTCTGGATTCA... 290 354037 IGHV3-43 01 IGHJ6 02 61.69 61.69

3 GCCTCTGGATTCA... 290 6798 IGHV3-43_01 IGHJ6_02 1.18 64.29

5 GCCTCTGGATTCA... 290 4082 IGHV3-43_01 IGHJ6_02 0.71 66.15

7 GCCTCTGGATTCC... 290 1672 IGHV3-43_01 IGHJ6_02 0.29 66.85

9 GCCTCTGGATTCA... 290 1361 IGHV3-43_01 IGHJ6_02 0.24 67.35

GCCTCTGGATTCA... 290 6602 IGHV3-43_01 IGHJ6_02 1.15

8 GCCTCTGGATTCA... 290 1494 IGHV3-43_01 IGHJ6_02 0.26

10 GCCTCTGGATTCA... 290 1314 IGHV3-43_01 IGHJ6_02 0.23

287 8147 IGHV3-43 01 IGHJ6 02 1.42

290 2364 IGHV3-43_01 IGHJ6_02 0.41

LymphoTrack[®] bioinformatic analysis begins on MiSeg[®] with stitching of paired reads (PGM[®] analysis begins at the next step). The resulting sequence file is then filtered for noisy reads (non-specific or fragments that are too short), and reads that do not pass quality are removed. These reads are then aligned to the reference sequence database, and numerous statistics are calculated. SHM rate is calculated from the alignable portion of the V-gene. MRD detection requires the previous identification of a clonal read, and for the user to input the amount of DNA being assaved. MRD results are reported for exact matches, and very similar matches to the clone submitted by the user. A confidence value is assigned to these results that tell the user the reliability of result, which is based on how much DNA was assayed, as well as the read depth achieved for that sample. The confidence is based on theoretical samplings using a binomial distribution.

63.10

65 44

66.56

67 11

67.58

5 24

5.24

5.68

5 68

5.24

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5.68

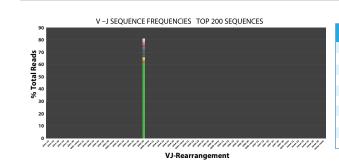


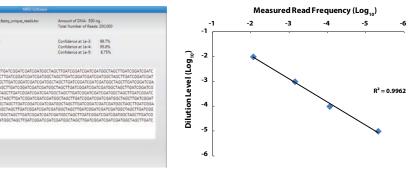
FIGURE 3. LYMPHOTRACK® SOFTWARE OUTPUTS

A typical MiSeq" IGH FR1 result from a bone marrow sample showing evidence for clonality with one sequence at 62% of total reads and mutation rate of 5%. Top 10 unmerged reads are presented in the table. FASTQ data from either MiSeq* or PGM* can be analyzed by LymphoTrack* Software running on a Windows* PC. The LymphoTrack* Software generates frequency distributions, DNA sequences, V-J assignment and usage, and SHM status. An add-on module allows for the tracking of clonotypes for MRD purposes.

GCCTCTGGATTCA

GCCTCTGGATTCA

Results



frequency, as well as confidence metrics for various levels of detection. Additional outputs will include sequence files for all reads

FIGURE 6. MRD DILUTION CONCORDANCE

Dilution experiment shows strong linear concordance between dilution level and measured read frequency.

Conclusions